

LeFever, H.M. Kansas State Teachers College, Emporia. Analysis of three White mutants resulting in two new recombination sites at the white locus in *Drosophila melanogaster*.

Three viable white alleles have been analyzed in respect to their location in the locus and their relationship to the mutant zeste. One of the mutants was white-carrot (w^{crr}) which arose spontaneously in a wild type X-chromosome (Judd 1964). The two remaining mutants were recovered by this author and designated as w^{65a25} and

$sp-w^4$. When tested with $sp-w^1$, the mutants w^{crr} and w^{65a25} gave the uniform brown eye color characteristic of a white allele rather than a white deficiency. The spotted white phenotype is relatively rare among white mutants and three previous mutants of this type were localized at what is now site 7 (Figure 1). Females with the genotype $sp-w^1/sp-w^4$ had a phenotype indistinguishable from females with the genotype $sp-w^4/sp-w^4$ or $sp-w^1/sp-w^1$ indicating that $sp-w^4$ was possibly a true allele of site 7.

	w^{Bwx}	w^{bf}	w^{crr}	w^{65a25}	w^a	w^1	$sp-w$
Judd 1964	0.01+		0.001+		0.01+	0.005+	
	1	2	3	4	5	6	7
LeFever 1973		0.0004+	0.0003+	0.002+		0.014+	

Figure 1. Map illustrating the seven recombination sites of the white locus. For descriptions of the mutants and symbols used, see Lindsley and Grell, 1968. The map distances are corrected for the presence of the autosomal inversions which increase the rate of crossing over by a factor of approximately 3X.

Recombination analysis was used to determine the spatial relationship of the three mutants with reference to the white locus. All constructed parental females carried the autosomal inversions $SMI/+$ and $Ubx^{130}/+$ which were employed to increase crossing over in the distal portion of the X-chromosome. Judd (1964) reported that w^{crr} recombined with w^a and was located to the left of it (Table 1). Females were constructed with the genotype $w^{crr}/y\ z\ w^{Bwx}$ and mated to $y\ w\ spl\ sn^3$ males. Ten exceptional progeny were recovered which place w^{crr} to the right of w^{Bwx} (Table 1). Females were constructed with the genotype $w^{crr}/y\ z\ w^{bf}$ and mated to $y^2\ w^a\ spl\ sn^3$. Two exceptional progeny were recovered which place w^{crr} to the right of w^{bf} (Table 1). Females were constructed with the genotype $z\ w^{65a25}\ spl\ sn^3/y^2\ w^a\ spl\ ec$ and mated to $y\ w\ spl\ sn^3$ males. One exceptional male progeny was recovered which places w^{65a25} to the left of w^a (Table 1). Females were constructed with the genotype $z\ w^{65a25}\ spl\ sn^3/w^{crr}$ and mated to $y\ w\ spl\ sn^3$ males. Two exceptional progeny were recovered which place w^{65a25} to the right of w^{crr} (Table 1). The mutant $sp-w^4$ was tested in the following manner: females were constructed with the genotype $y^2\ sp-w^4\ spl\ sn^3/y^2\ w^a\ spl\ ec$ and mated to $y\ w\ spl\ sn^3$ males. Nine exceptional progeny were recovered which place the mutant $sp-w^4$ to the right of w^a (Table 1).

When the mutants w^{crr} and w^{65a25} were placed in the heterozygous state with two doses of zeste ($z\ w^+ \text{dup ec}/w^{crr}$ and $z/z\ w^{65a25}\ spl\ sn^3$) the results were a female with a zeste phenotype. This indicated that w^{crr} and w^{65a25} do not affect the expression of zeste. The mutant $sp-w^4$, according to its placement by recombination, should be a zeste suppressor (Green 1959c). This was the case as females with the genotype $y^2\ sp-w^4\ spl\ sn^3/z\ w^+ \text{dup ec}$ had a nearly wild type phenotype characteristic of a zeste suppressor effect.

Analysis of the mutant w^{crr} and w^{65a25} which places them in separate locations between former recombinational site 2 (w^{bf}) and site 3 (w^a) indicated that the white locus has at least 7 recombination sites of which sites 1 through 5 are non-suppressors of the mutant zeste (Figure 1).

Acknowledgments: I wish to thank Dr. Burke H. Judd for his assistance during the early part of this study, undertaken while I was a graduate student at the University of Texas, and

(Continued at bottom of next page)

Van Valen, L. University of Chicago, Illinois. A method that might estimate age in *Drosophila*.

In a previous note (DIS 46:125) I reported failure in an attempt to estimate age in *Drosophila* by use of daily growth layers in the cuticle. Recently Schlein and Gratz (1972) have had success with this method for mosquitoes

and three families of muscoid flies. Their methods differ from ours in perhaps relevant ways. Success with *Drosophila* could make its ecology amenable to standard ecological procedures. I now lack the relevant equipment but suggest that my previous failure not be taken as definitive.

Reference: Schlein, J. and N.G. Gratz 1972, Bull. World Health Org. 47:71-75.

(Continued from preceding page)

Table 1. Exceptional recombinant types recovered from heterozygous females.

Heterozygous female	Exceptional recombinant types recovered	Number	Total offspring
$y^2 + w^a spl ec$ **	$+ w^{crr} w^a spl ec$	4	52713
$+ w^{crr} + + +$	$y^2 + + +$	1	
$+ + + w^{crr}$	$y z w^{Bwx} w^{crr}$	3	103586
$y z w^{Bwx} +$	$z w^{Bwx} w^{crr}$	1	
	$+ + + +$	6	
$+ + + +^{crr}$	$+ + + +$	2	170725
$y z w^{bf} +$			
$+ Z w^{65a25} + spl + sn^3$	$y^2 + spl sn^3$	1	14506
$y + + w^a spl ec +$			
$z + w^{65a25} spl sn^3$	$z + + + +$	2	255341
$+ w^{crr} + + +$			
$y^2 + sp-w^4 spl + sn^3$	$y^2 + + spl ec$	5	21550
$y^2 w^a + spl ec +$	$y^2 w^a sp-w^4 spl sn^3$	4	

** Burke H. Judd personal communication

for his continued support and encouragement on this project.

References: Green, M.M. 1959c, Hered. 13:302-315; Judd, B.H. 1964, Genetics 49:253-265.